

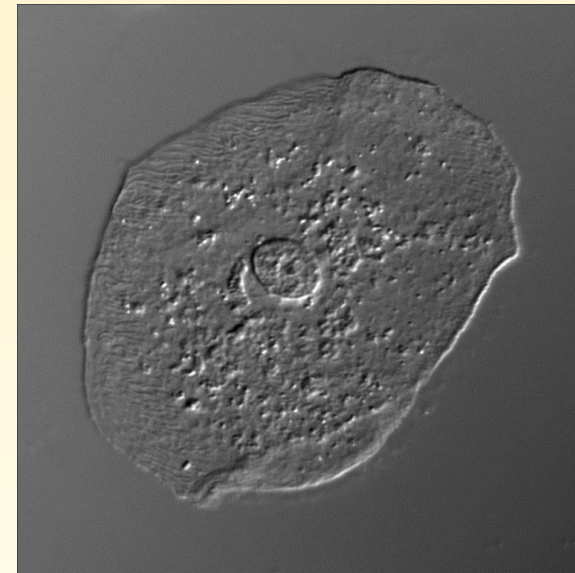
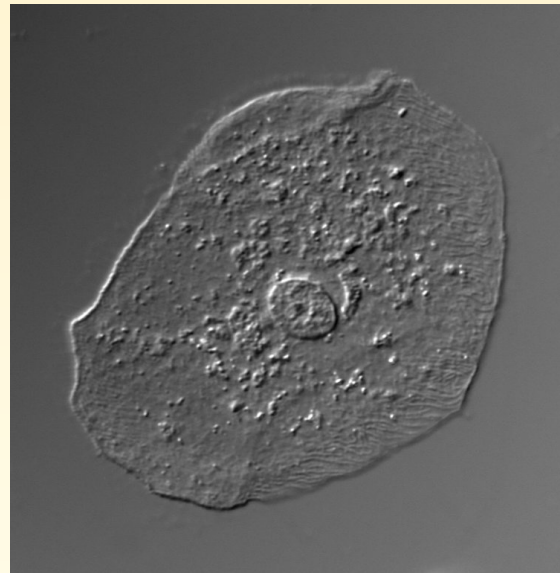
5. Differential Interference Contrast



Bright-field



Differential Interference Contrast (right: image turned)



cheek cells, as prepared in the RegBioMed course

5. 3D-effect done in the computer



Bright-field – plus contrast inverted image - overlay

Bright-field



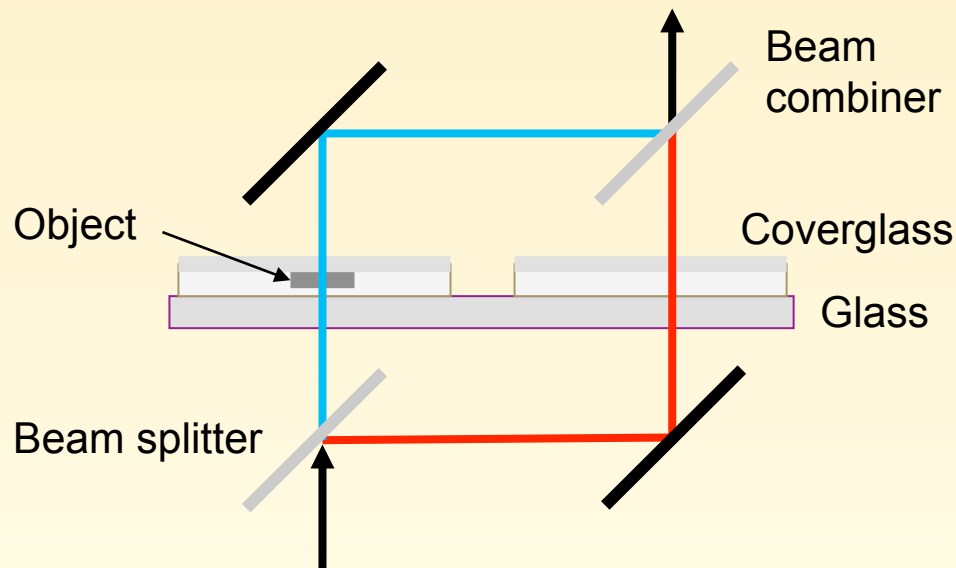
cheek cells, as prepared in the RegBioMed course



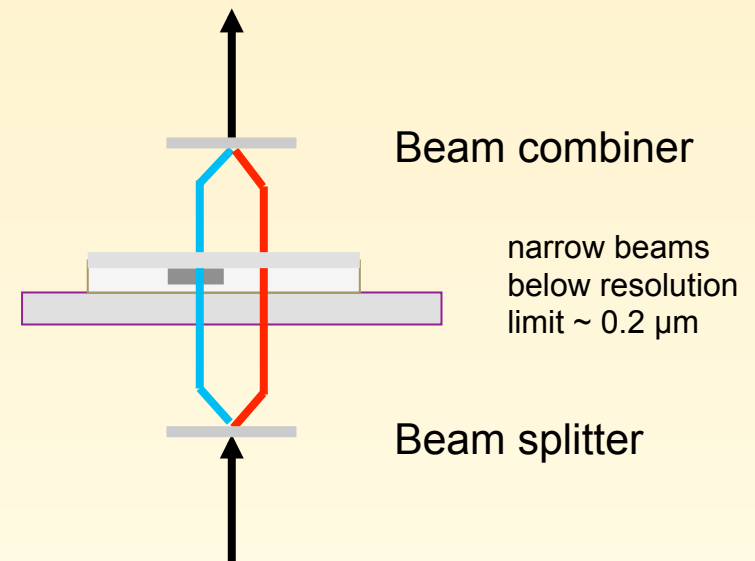
5. DIC - interferometry



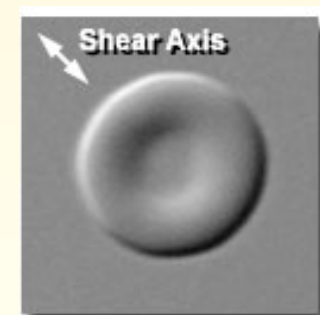
Classical double-path interferometer with a real reference sample



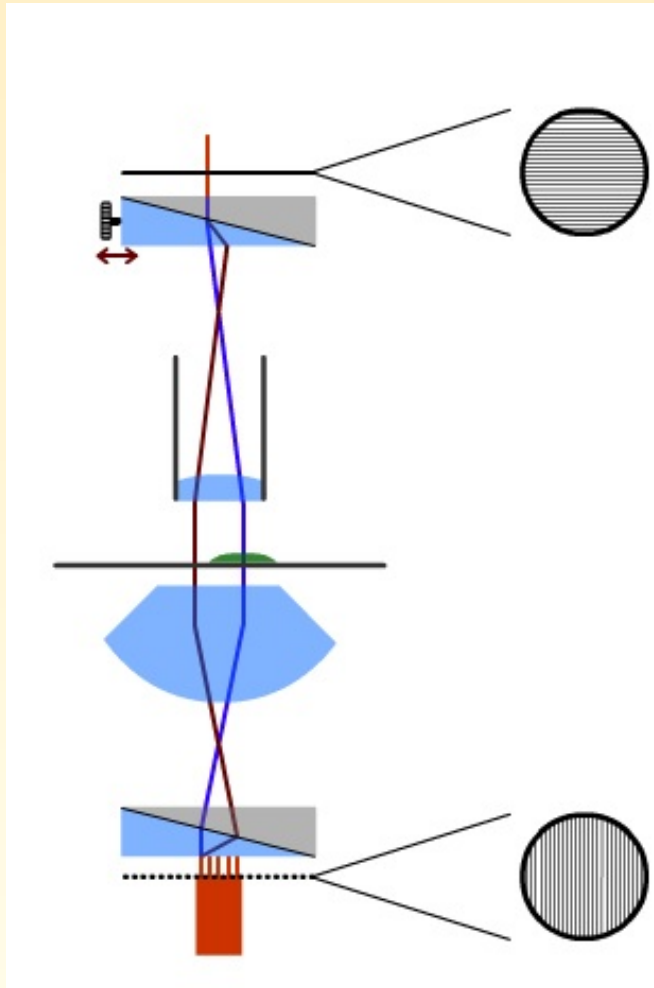
Used for DIC:
Common path interferometer,
lateral shearing interferometer



- superimposed waves interfere
- information about optical path difference (caused by Δx or Δn)
- DIC: contrast is proportional to the path length gradient (i.e. differential) along the shear direction (i.e. edge contrast)



5. DIC – optical setup overview



analyzer: beams interfere

objective prism reunites the beams

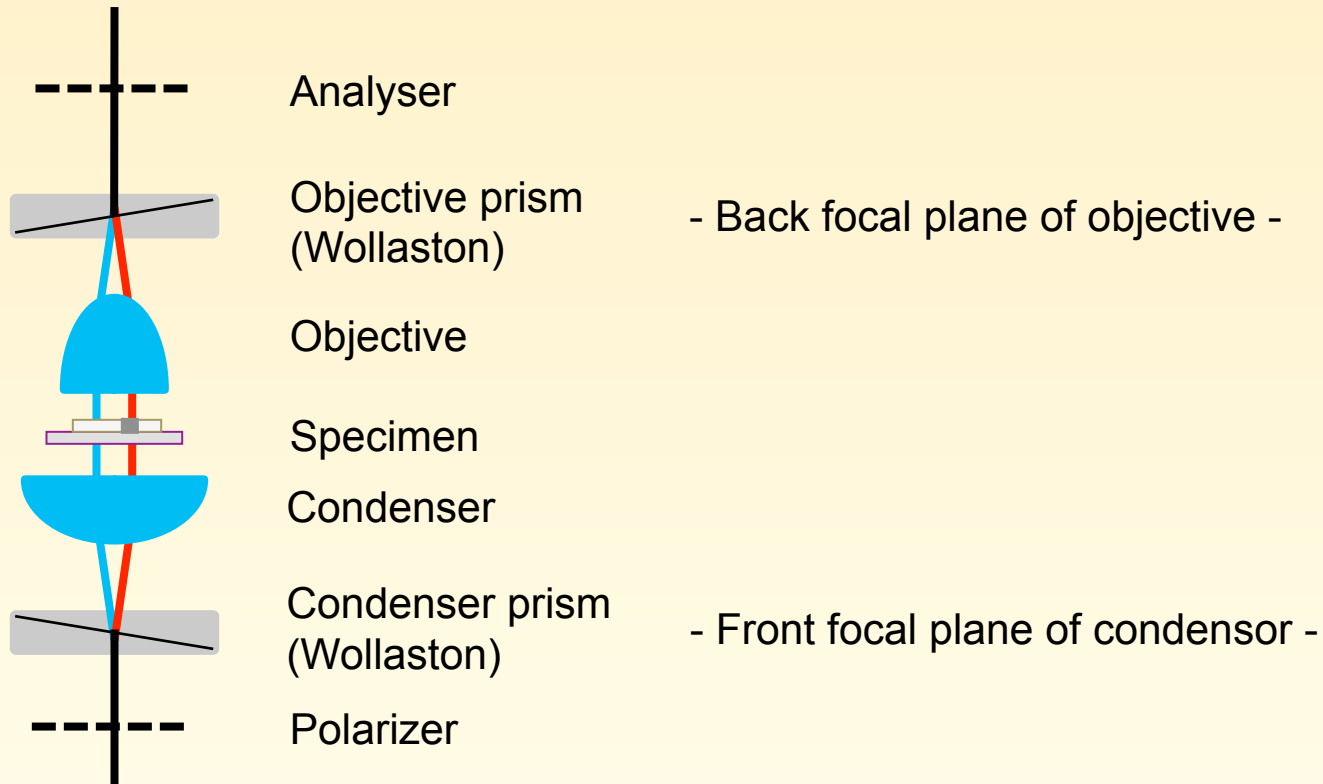
prism can be shifted horizontally to tune phase shift

two narrow beams below resolution limit: phase shifts at edges of sample or structures

condenser prism: splits in two beams, mutually orthogonal linearly polarized

polarizer: creates linearly polarized light

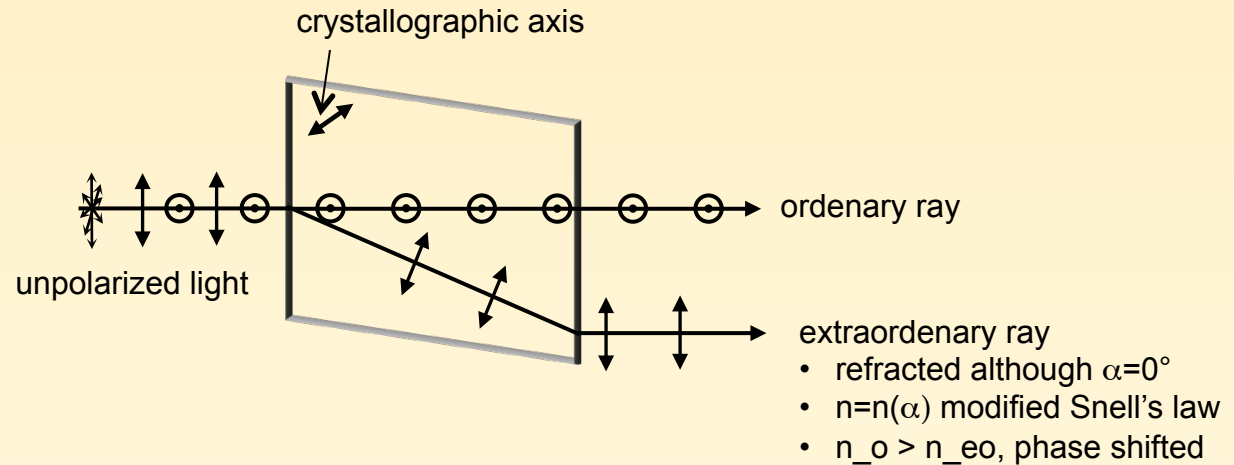
5. DIC – beam splitting, where?



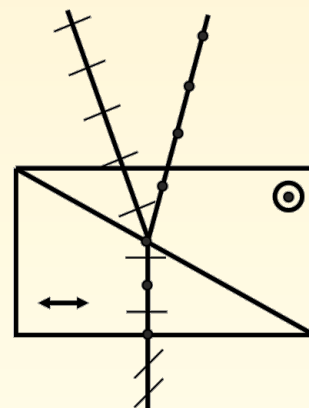
5. DIC – beam splitting, how?



Calcite (CaCO_3) crystal:
double refractive material



Condensor prism
(type Wollaston)



45° linearly polarized

- made out of two glued calcite wedges
- perpendicular crystallographic axes
- axis of first prism 90° to beam direction
- 45° linearly polarized incoming beam

5. DIC – Smith vs. Nomarski



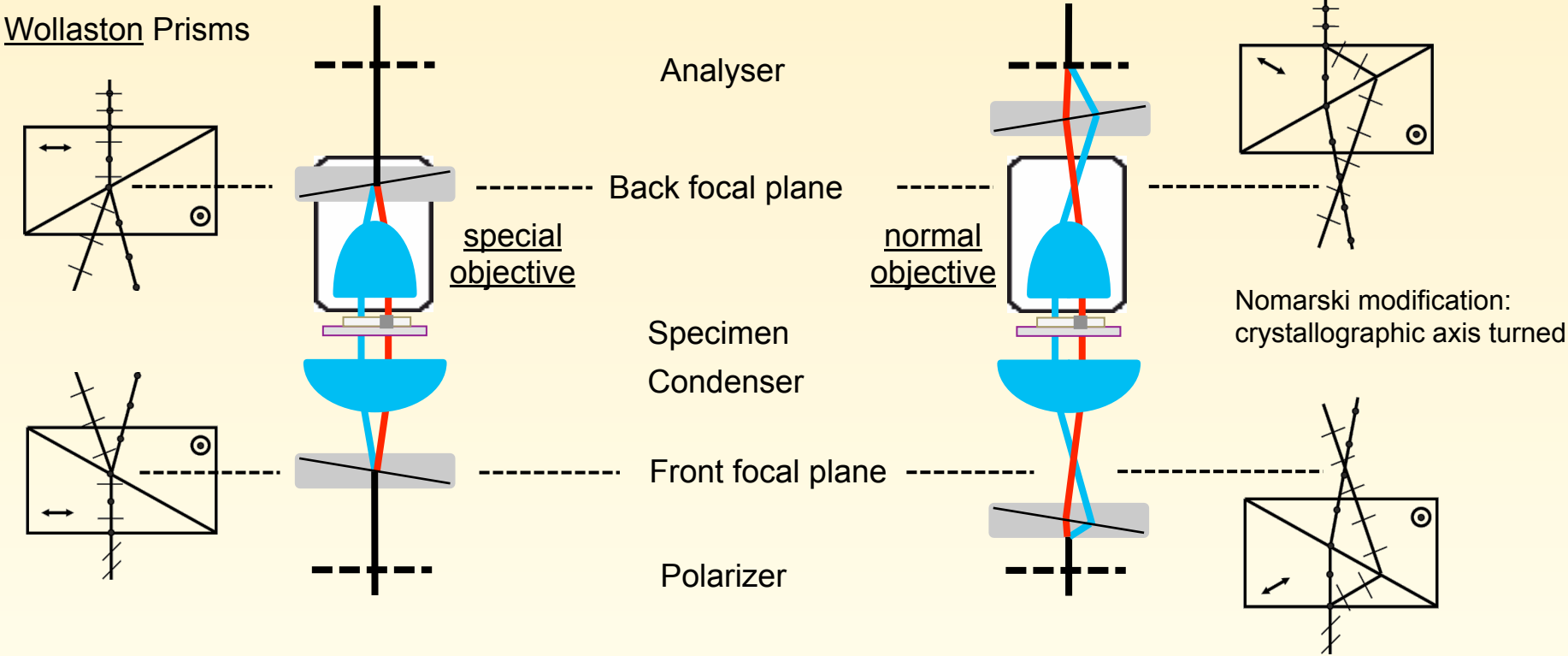
Interference microscope after ...

SMITH

NOMARSKI

Wollaston Prisms

Nomarski Prisms

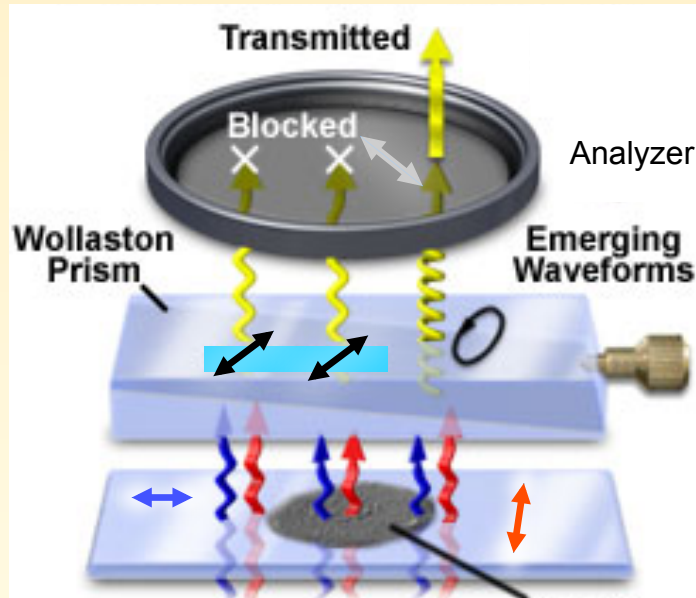


→ preferred configuration

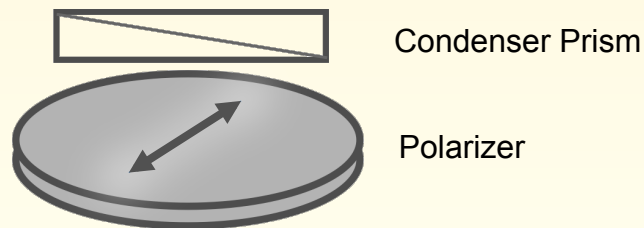
5. DIC – phase shifts



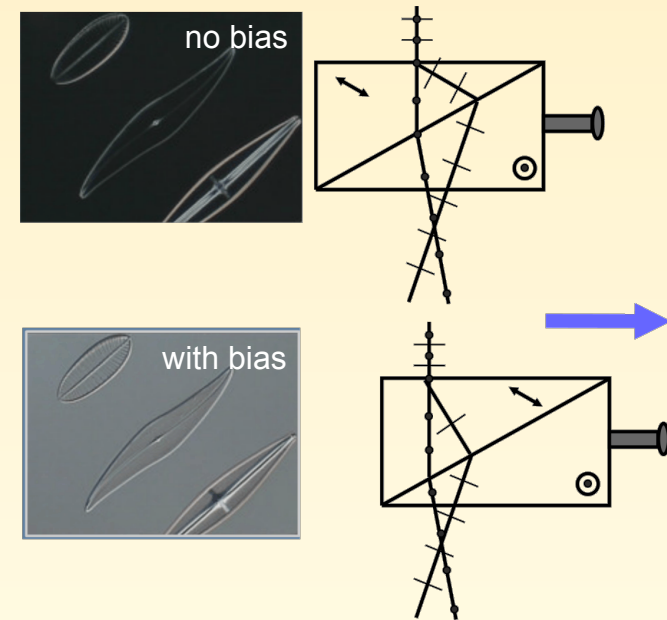
by the phase SAMPLE



phase specimen: phase shift between both beams



by BIAS retardation



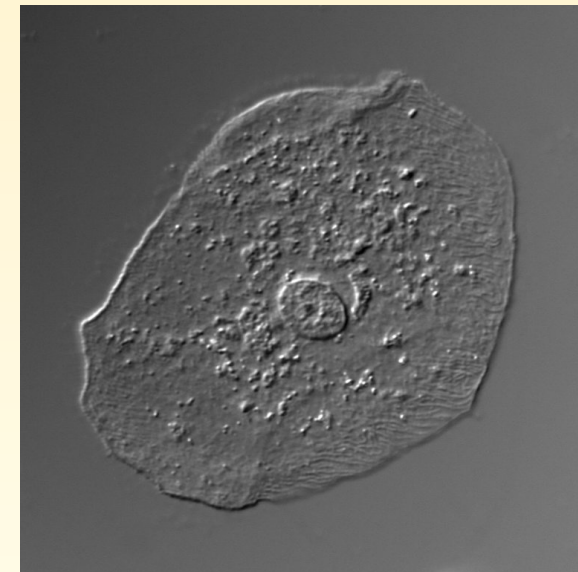
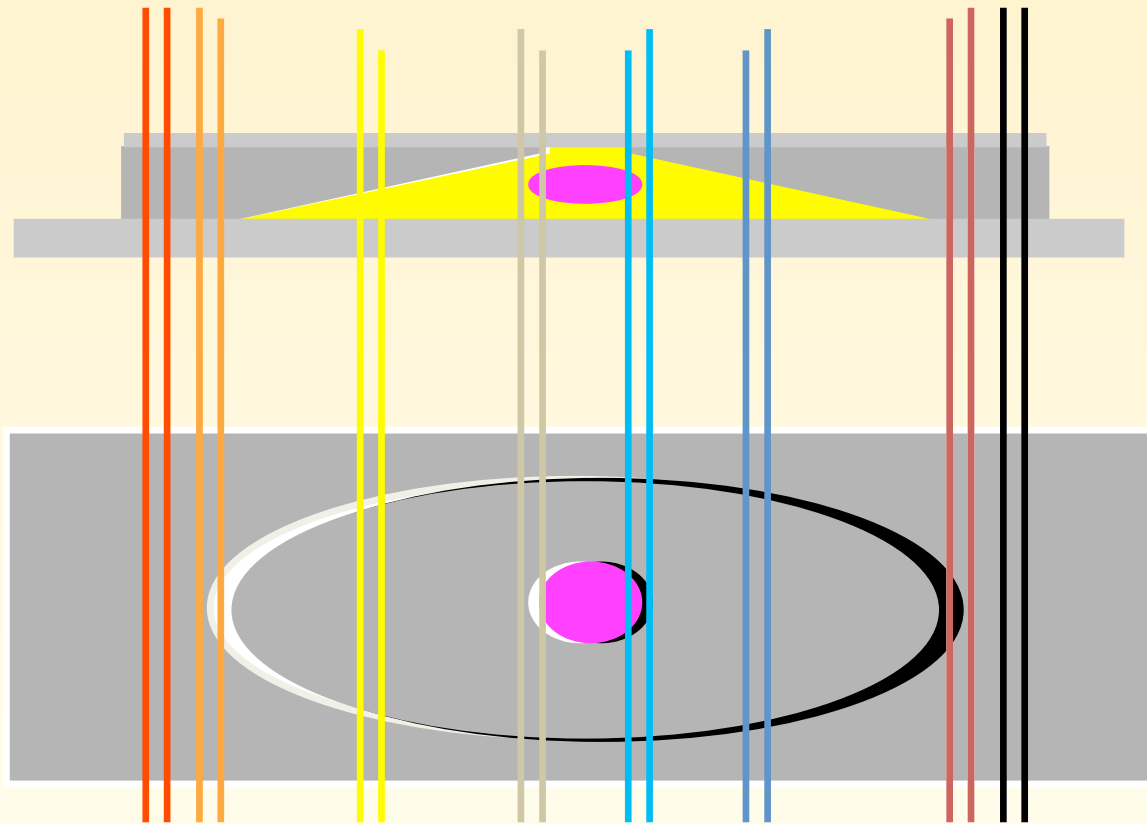
- lateral shift of the objective prism (Nomarski)
- bias, i.e. offset phase shift, grey background
- features bright and dark on different sides

5. DIC – phase shifts in the sample



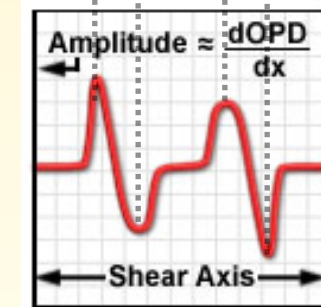
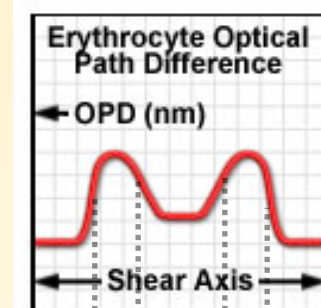
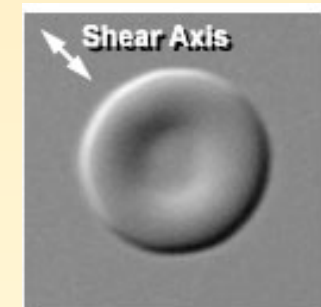
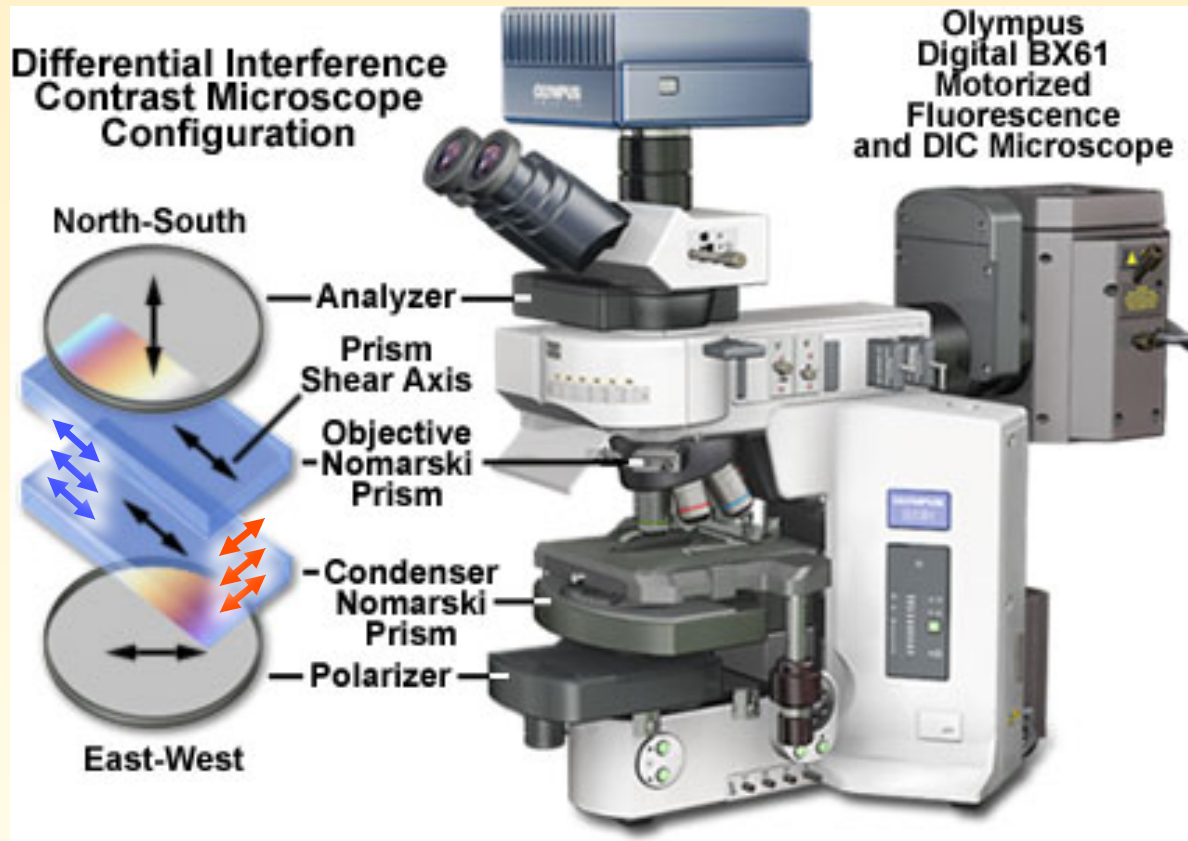
... for example a cell: phase shifts occur at:

- edges of the cell
- edges of subcellular components



cheek cell

5. DIC – microscope setup



$d OPD / dx$ is the differential of the optical path difference (thickness or Δn) along the shear axis x

5. Differential Interference Contrast



Try yourself

Setup:

- Köhler the microscope
- adjust polarizer and analyzer – crossed polars (observe BFP as well)
- put in the correct condensor prism (observe BFP as well)
- put in correct objective prism, first separately, then both prisms
- tune the objective prism for the best DIC effect (best “shadows”, colors)

Specimen:

- diatoms (observe which substructures give strongest effect, then turn the sample)
- cheek cells

Additional information:

- no plastic dishes, as it is birefringent. Only use glass, or glass bottom dishes (no lid).
- some objectives are especially suited for polarisation and DIC. They are labeled with “DIC” or labels are in red letters. All other objectives still work.

DIC Example

