Light Microscopy course Differential Interference Contrast DIC











DIC – differential interference contrast

 developed in the mid-1950s by Georges (Jerzy) Nomarski (1919-1997), a Polish optics theoretician working in France at CNRS

For a detailed biography see: http://micro.magnet.fsu.edu/ optics/timeline/people/ nomarski.html







DIC – probe **differential** by interference = contrast

- Differential = (minute) difference between two different values of something (given in Δ or d) over a certain (small) range (gradient or slope = $\Delta y/\Delta x$)
- Used in many different contexts (Math, Physics, Engineering, Biology....)





DIC – probe **differential** by interference = contrast – example for differential





DIC – probe **differential** by interference = contrast – example for differential



DIC – probe **differential** by interference = contrast – example for differential





DIC – differential by **interference** = contrast

Interference is a phenomenon in which two waves superpose to form a resultant wave of greater, lower or same amplitude. (Wikipedia)



Wikipedia



https://machinesdontcare.wordpress.com/2011/02/05/ interference-patterns/interference-fringes-04/







DIC – differential by **interference** = contrast The principle of DIC - interferometry

Classical double-path interferometer with a real reference sample











DIC – differential by **interference** = contrast





30 degrees slower





(q)

(a)

DIC – differential by **interference** = contrast (q 1.5 (a) 1 0.5 0 120 180 240 300 360 420 480 540 600 660 720 60 -0.5 -1 -1.5









150 degrees slower











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The principle of DIC - interferometry

Common path interferometer, lateral shearing interferometer









DIC – differential by interference = contrast

Differential Interference Contrast Imaging of Transparent Thin Specimens



From ZEISS-campus website







DIC – introducing retardation bias







$\lambda/4$ bias + further $\lambda/6$ retardation of blue beam







$\lambda/4$ bias of blue + $\lambda/6$ retardation of red beam









With $\lambda/4$ bias of blue beam



Resultants of retardation of beam by 60 deg ($\lambda/6$)



grey: no retardation, dark: blue retarded, light: red retarded





Interference at analyzer







Without bias $\lambda/2$ shift at analyser With $\lambda/4$ bias





Resultants of retardation of beam by 60 deg ($\lambda/6$)



grey: no retardation, dark: blue retarded, light: red retarded



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Back focal plane of objective

Objective Specimen

Condenser

Front focal plane of condenser

adapted from Peter Evennett











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The Wollaston prism – how does it work

- Remember birefringence in a Calcite crystal:



- two spots out of one
- But: one looks to be further away (ie had a retardation introduced in its path)







The Wollaston prism beam splitter – how does it work

direction of polarisation



Shear = spatial separation of the two beams



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The Wollaston prism – beam combiner

direction of polarisation

Wikipedia

orientation of analyser



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Problem with objective Wollaston prism



Analyser

Back focal plane of objective

Objective Specimen

Condenser

Front focal plane of condenser

Polariser

adapted from Peter Evennett







Wollaston versus Nomarski-modified prism







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Examples of DIC – what it is good for



axis of shear

Unlabeled human RBCs in buffer on uncoated glass cover slip. Zeiss Axiovert 200M, 100x / 1.4 oil DIC.



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Examples of DIC – what it is good for



axis of shear

Zebrafish keratocytes

speed of cells: ~ 13.5um/min; 3min 30sec movie

Zeiss Axiovert 200M, 100x 1.4 oil DIC





Examples of DIC – what it is good for



C. elegans embryos Gunar Fabig, MTZ





Examples of DIC - summary

- Good for thin or thicker specimen
- labeled or unlabeled specimen
- Highlights gradients in optical path differences, not absolute optical path values
- For contrast in DIC shape of an object is more important than the absolute phase shift produced by the specimen









Examples of DIC – limitations

- Samples must be placed in an non-birefringent environment (no plastic dishes, no plastic lids!)
- dependent on perfectly set up Koehler illumination and strain free optics
- some objectives are especially suited for polarization and DIC:
 - labeled with "DIC" or labels in red letters.
 - all other objectives still work too, but there might be quality problems
- contrast only achieved in direction of shear between the two beams!







1) Köhler your microscope carefully!











2) Insert and adjust polarizer and analyzer – crossed polars









3) If microscope has a De Senarmont compensator, put it into its position of zero bias







4) Put in correct objective prism into back focal plane (BFP) - related position of objective







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- 5) Swing in correct condenser prism in front focal plane
 - Roman number (I, II, or III) has to correspond to the number on objective prism used









6) check BFP (using Betrand lens or telescope) for image of blurred cross or move objective prism into position were background is darkest (maximum extinction)



from olympusmicro.com







turn objective prism or compensator for optimal bias





7)

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Use either cheek cells or diatoms





from photomacrography.net





