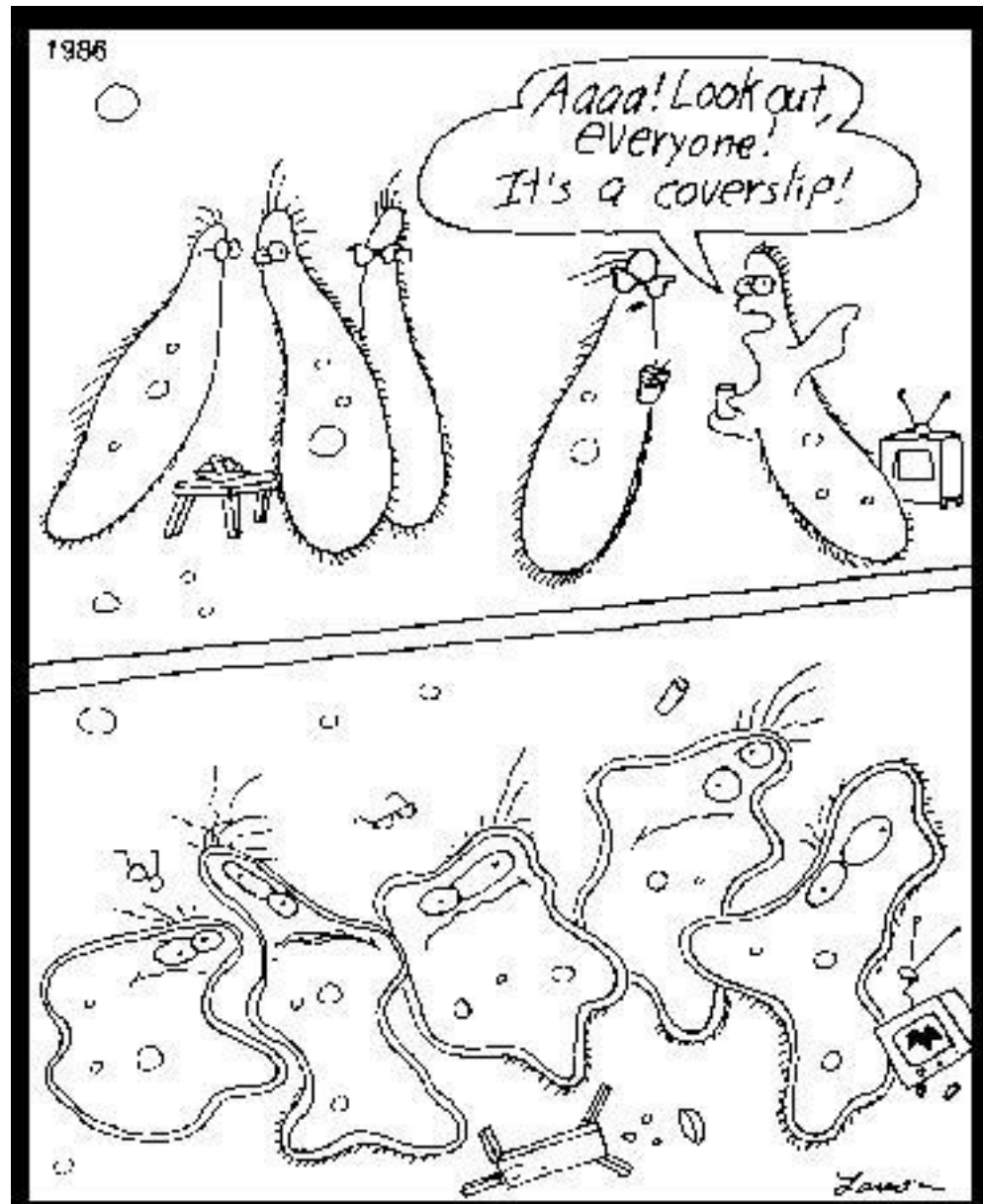


Selective plane illumination  
microscopy – SPIM  
(light sheet microscopy)

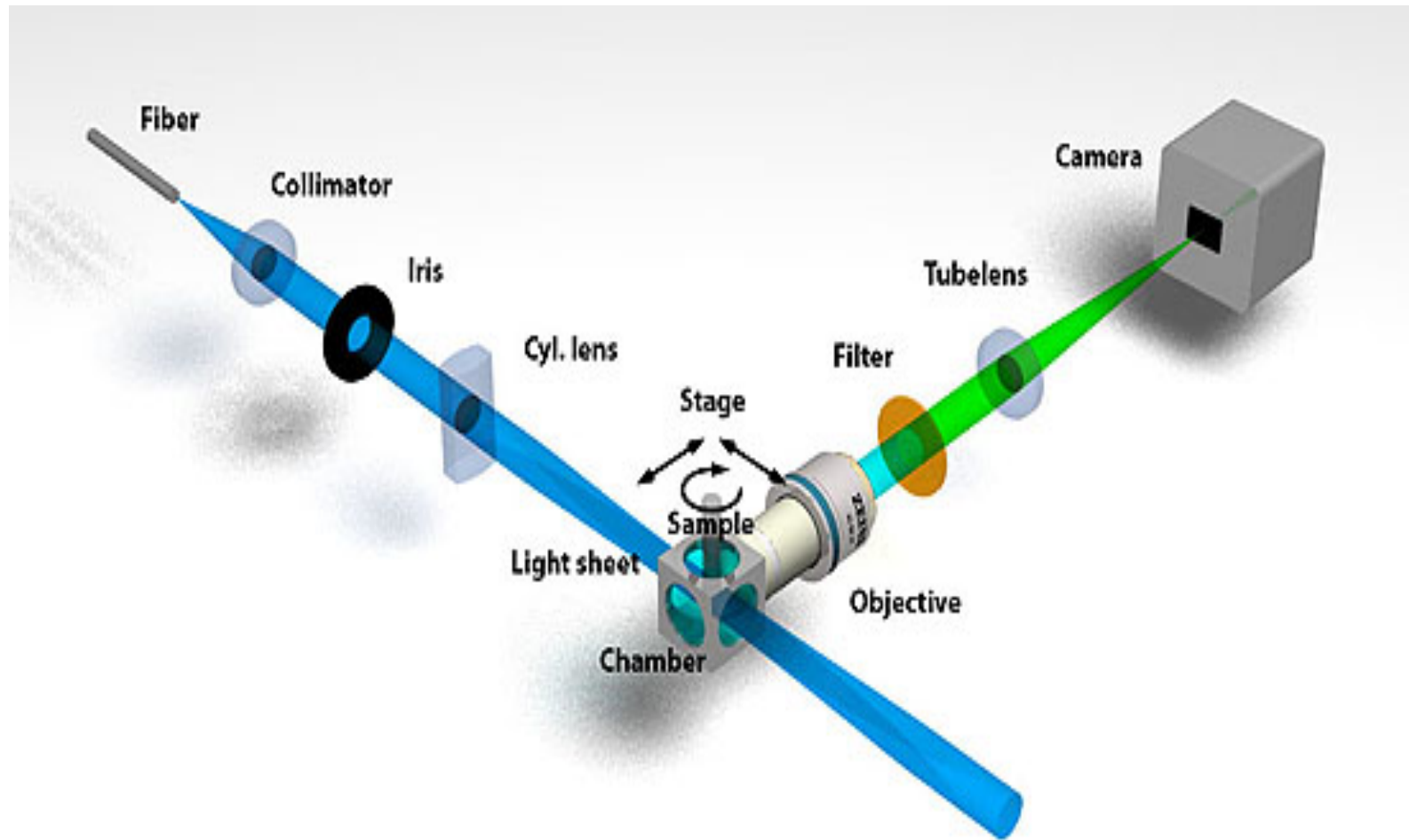
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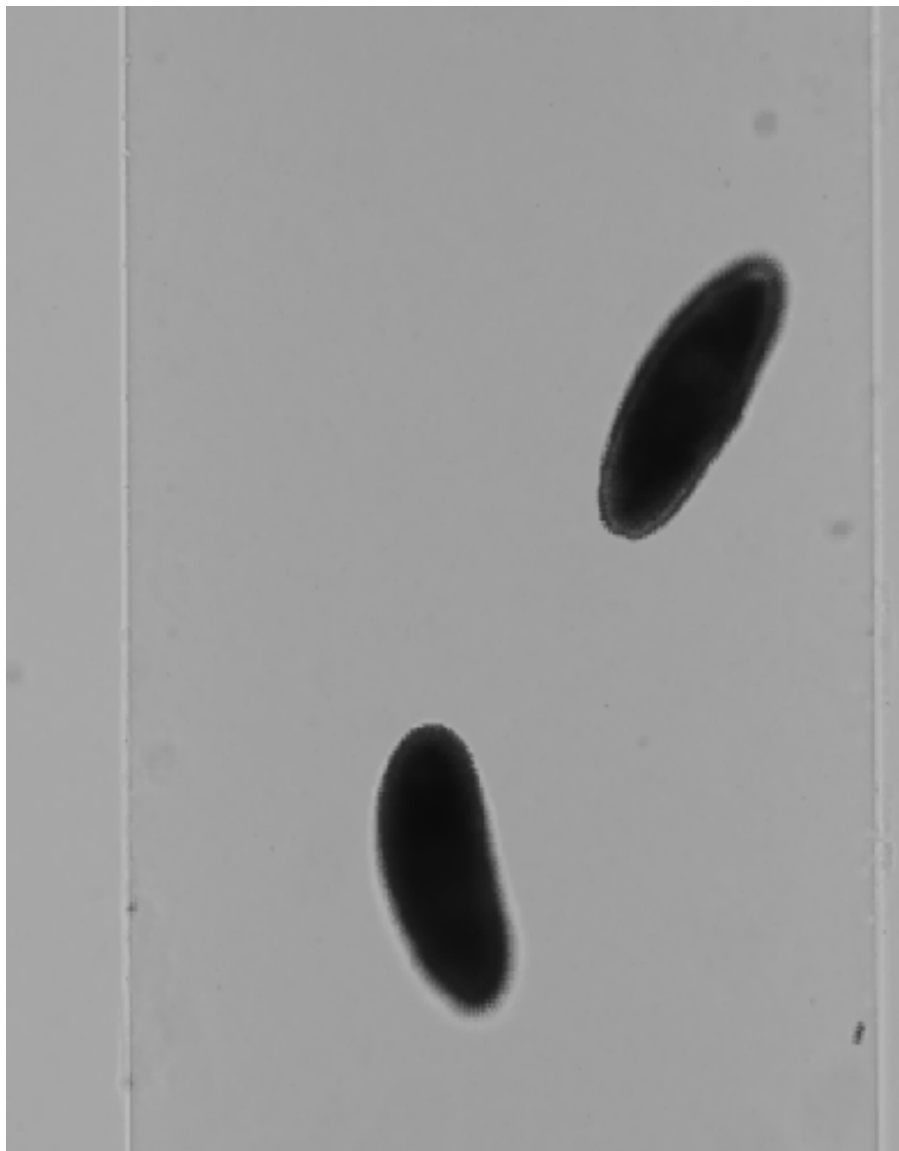
Life on a microscope slide

**Sticking biology to  
flat glass is not  
very  
physiological...**

# SPIM



<http://www.huisken.org/jan/spim.html>



Pavel Tomancak

# Selective plane illumination microscopy - SPIM

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Field illuminating technique (detector: CCD, EM CCD, sCMOS camera)

Temporal resolution typically tens to hundreds of ms/image in “standard” FOV – 512x512 pixels

Multiple angle image acquisition – isotropic resolution x,y,z

Low photobleaching

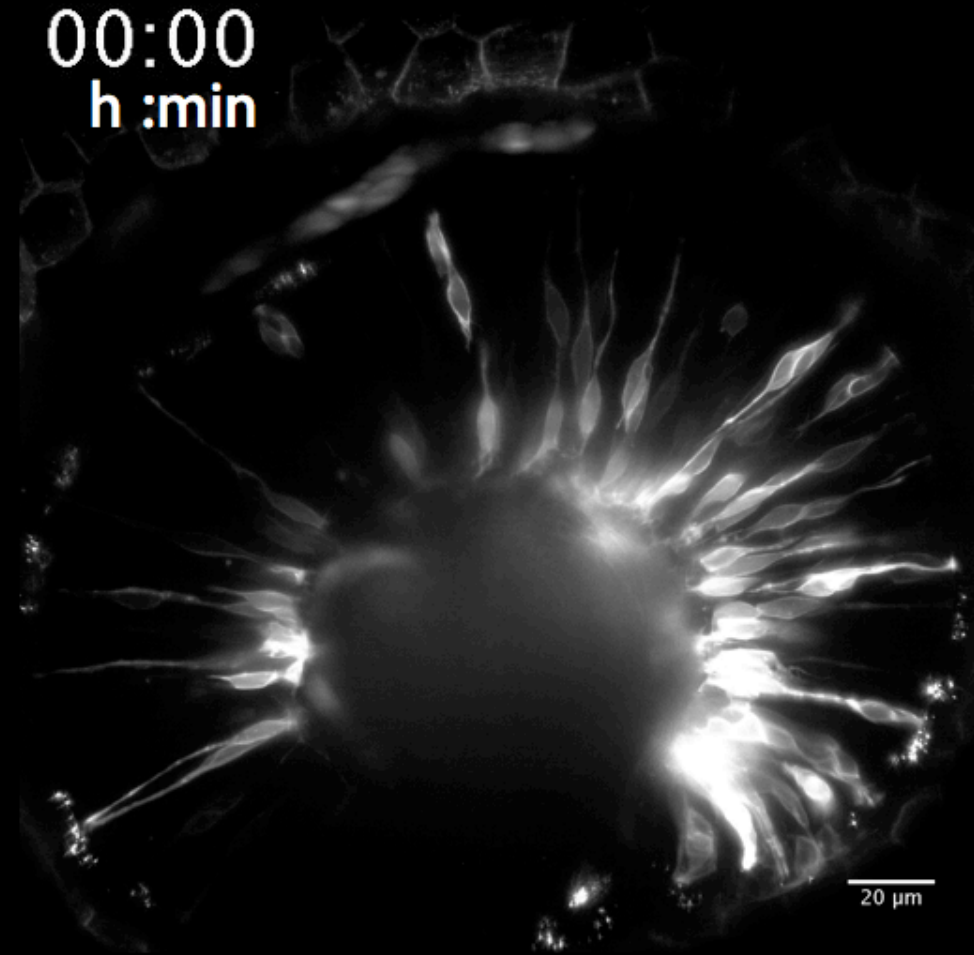
Compromise between size of FOV and resolution

# SPIM – what is it good for ?

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- live cell imaging of embryos
- multiple angle imaging of fixed and living specimens

# Neurogenesis in *Danio rerio* retina:

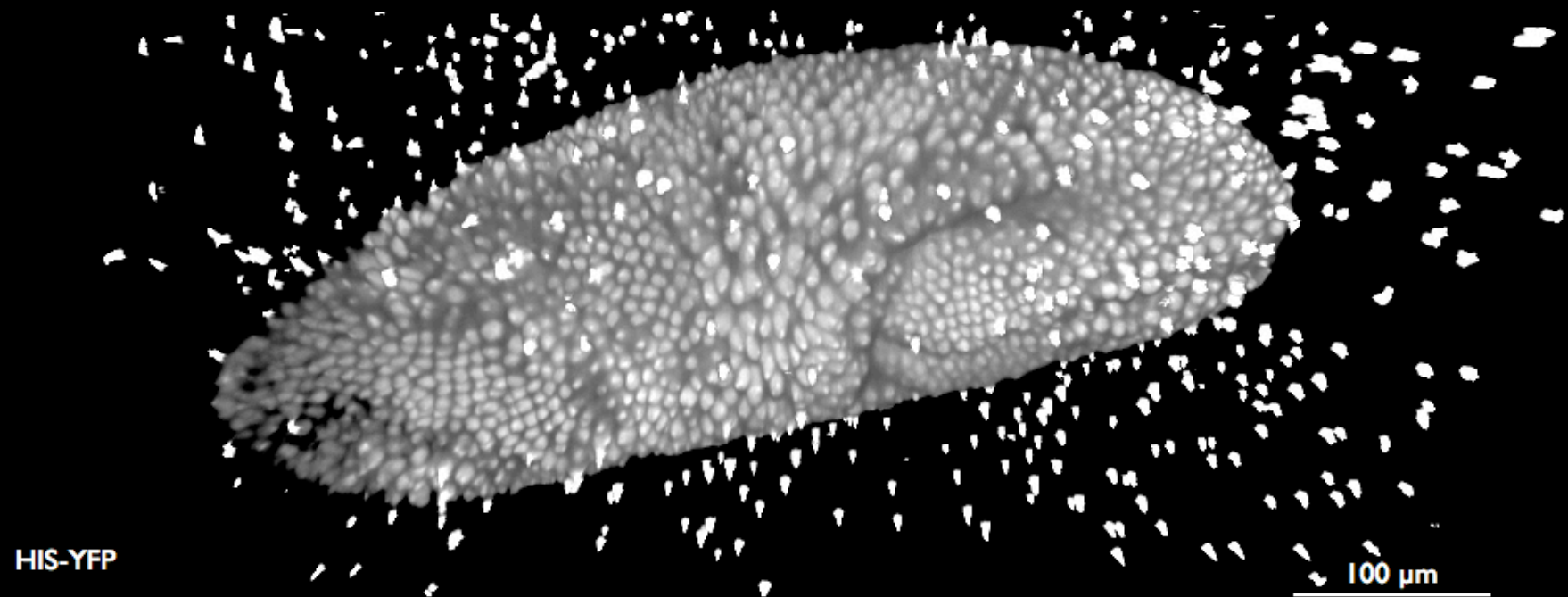


**Ath5-RFP**

Jaroslav Icha, Norden Lab, MPI-CBG

# Light-sheet microscopy:

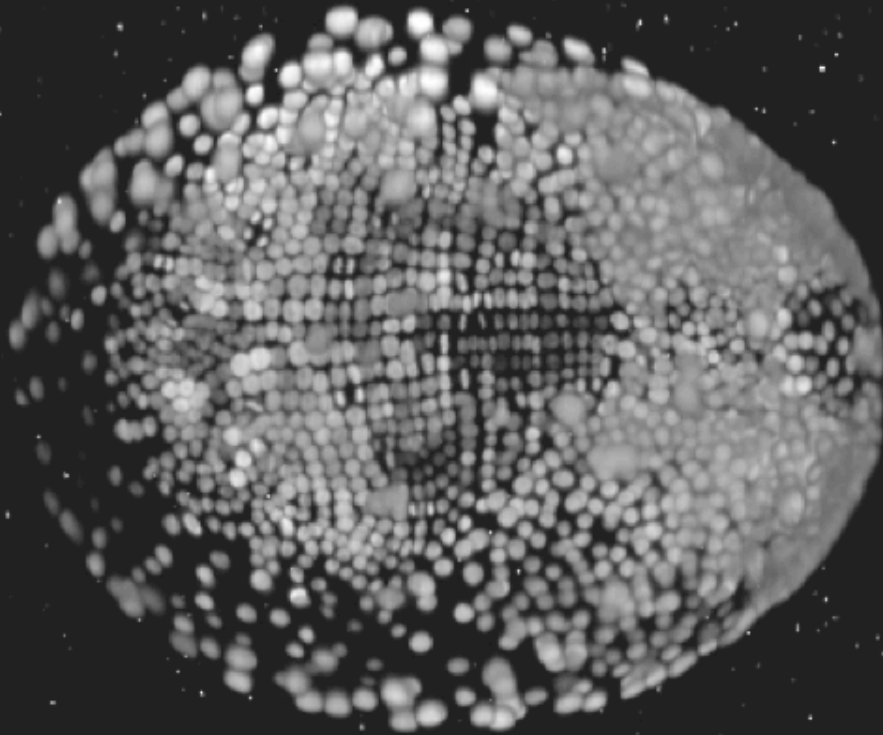
03 h : 31 min



Fly embryo development  
Valia Stamatakis, Tomancak lab - now  
postdoc in Janelia Farm (US)



0 d : 01 hours



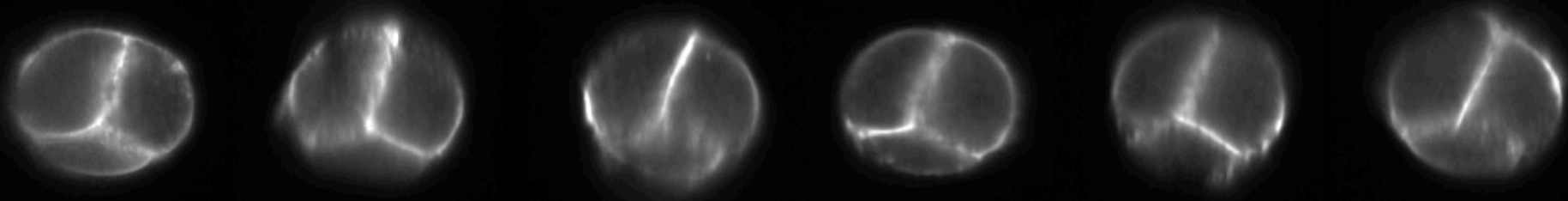
Tassos  
Pavlopoulos



*Parhyale hawaiiensis* HisRFP 888 time-points, 3-5 dual sided illumination angles, every 7.5 minutes, acquired on Lightsheet Z1, registered, fused and rendered in Fiji, 5.1 TB raw data

# Multi-view Fusion

*C. Elegans* embryo  
4-cell stage  
Ph-GFP Lipid binding domain



Angle 0

Angle 60

Angle 120

Angle 180

Angle 240

Angle 300

**Content based Fusion**



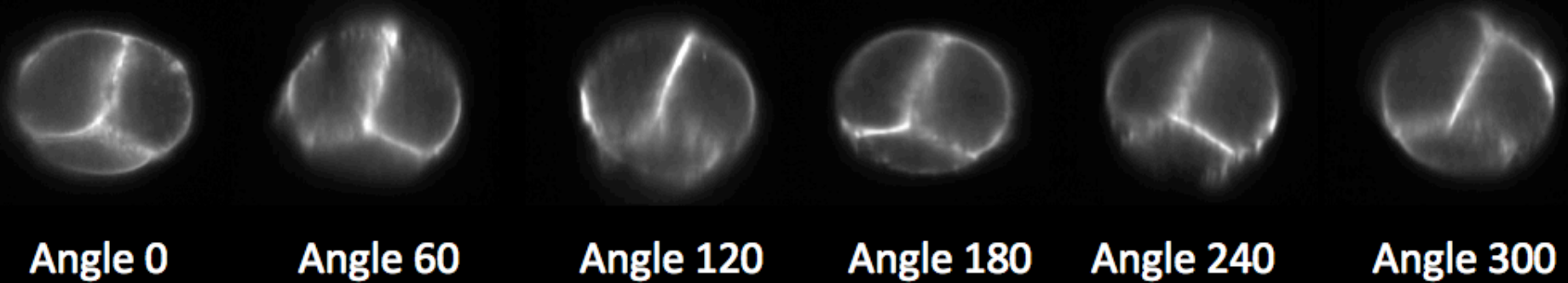
xy

xz

yz

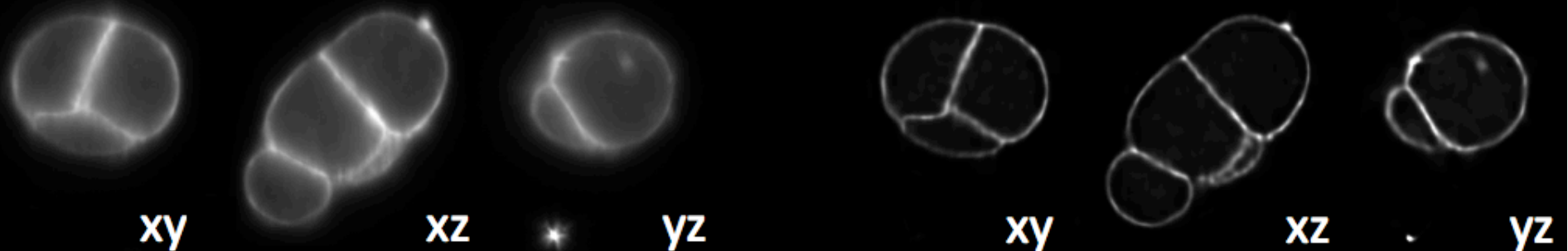
# Multi-View Deconvolution

*C. Elegans* embryo  
4-cell stage  
Ph-GFP Lipid binding domain

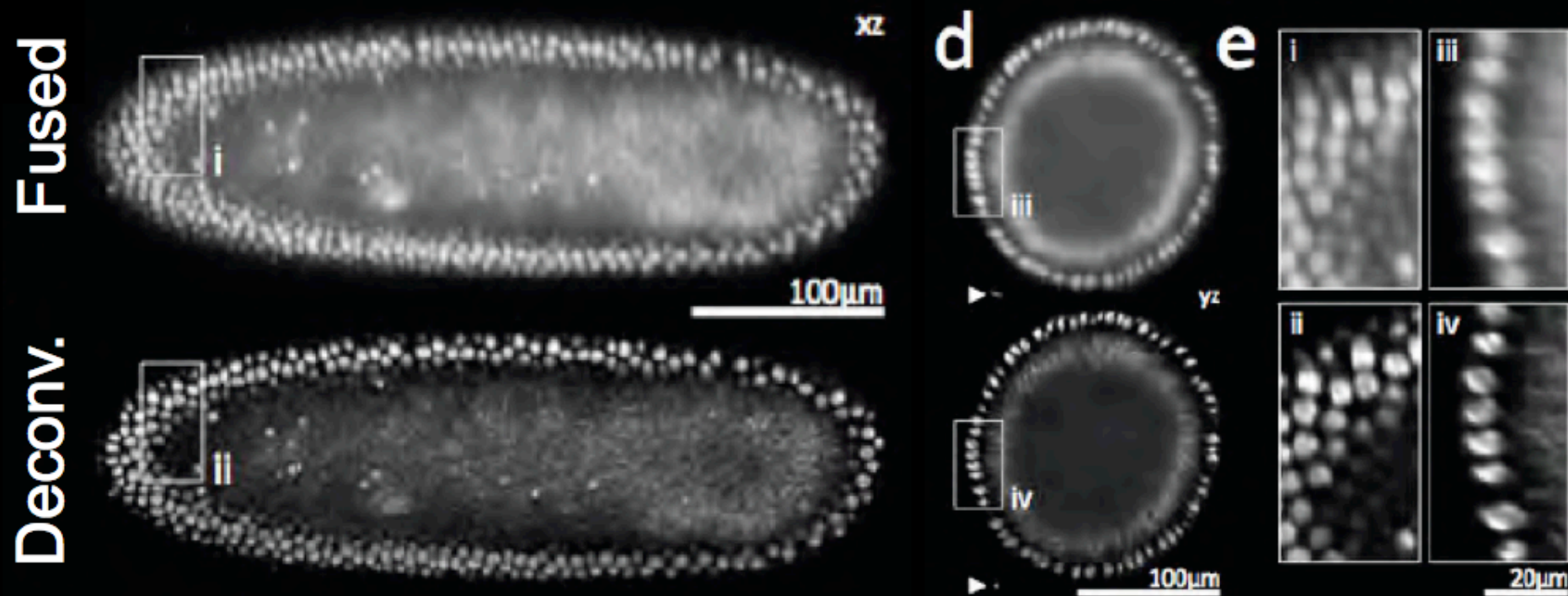


**Content based Fusion**

**Multi-View Deconvolution**



# Deconvolution results - Drosophila



His-YFP in all cells

# Deconvolution results - C. elegans

xy

yz

xz

nuclear envelope  
marker

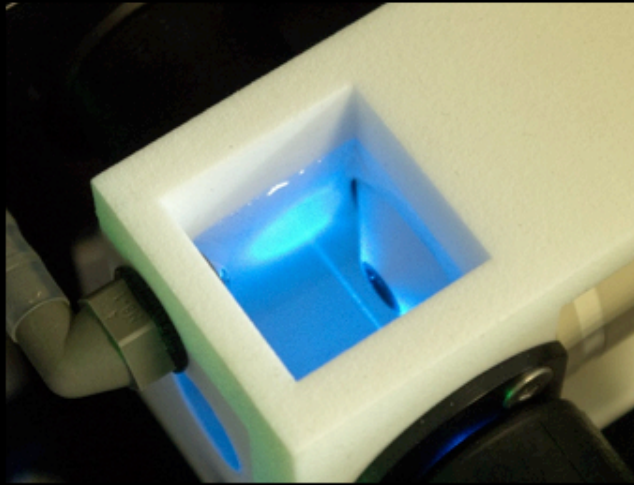
DAPI

# Problems of using SPIM technology:

Data storage

Data Transfer

Data processing



Data Analysis

3D visualization